FORMATION OF 30,120,150-TRIHYDROXYCHOLANIC ACID FROM DEOXYCHOLIC ACID BY FERRO-ASCORBATE SYSTEM

M. Kimura, M. Kawata, M. Tohma, A. Fujino and K. Yamasaki Faculty of Pharmaceutical Sciences, Hokkaido University Sapporo, Japan

(Received in Japan 6 April 1970; received in UK for publication 21 April 1970)

In 1965, B. Matkovics et al. 1 reported the in vitro transformation of deoxycholic acid (3 α , 12 α -dihydroxy-5 β -cholanic acid, I) into cholic acid (3 α ,7 α ,12 α -trihydroxy-5 β -cholanic acid, II) with ferro-ascorbate system containing ethylenediaminetetraacetic acid (EDTA). We have now succeeded in obtaining the trihydroxycholanic acid (III, m.p. 322-325° (decomp.)), instead of II as well as pythocholic acid (3 α ,12 α ,16 α -trihydroxy-5 β -cholanic acid, IV, m.p. 186-187°; methyl ester: m.p. 146-148°), from the reactions of I using the same systems both with and without EDTA in absolutely aqueous solutions.

Into the solution of I (1.0 g., 2.55 m moles), $FeSO_4 \cdot 7H_2O$ (5.0 g., 18.0 m moles) and ascorbic acid (100 g., 567 m moles) in 0.4 M phosphate buffer (pH 6.8, 2500 ml.), oxygen was bubbled for 3 hrs. at $37^{\circ}C$. Another 100 g. of ascorbic acid was then added to the reaction mixture which was adjusted again at pH 6.8 and kept at the same conditions as above for another 3 hrs. The products were extracted at pH 3.0 with ether and the chloroform-soluble substances from the ether extracts were then methylated. The methyl esters obtained were submitted to chromatography on silica gel by eluting with the mixture of benzene and ethyl acetate (EtOAc). The eluates by 70% EtOAc-benzene and 100% EtOAc gave the colourless needles (V): yield 3.3%; m.p. 256-259 $^{\circ}$ (EtOAc-MeOH); Calcd. for $C_{25}H_{42}O_5$: C 71.06 H 10.01, Found: C 71.35 H 10.15.

The fragmentation pattern of mass spectrum (Fig. 1) was considerably similar to that of methyl cholate: m/e 386 (M-2x18), 368 (M-3x18), 271 (M-(2x18+115), base peak], 253 {M-(3x18+115)}; 115 mass units may correspond to the intact side chain. The presence of three nuclear oxygen substituents may thus be elucidated. Chromate oxidation of V in acetic acid gave a triketo ester (VI); m.p. $176-177^{\circ}$, $^{\circ}_{CHC1}$, $^{\circ$

(3) can not be operative in 16-ketosteroid. The new oxygen function introduced in this hydroxylation reaction may, therefore, be situated at C_{15} .

In NMR studies on steroids, signal shifts of the angular methyl groups due to the hydroxyl groups in the various positions have been reported. The chemical shifts in six known bile acids as well as V and in some of those reported are shown in Table I. Contrary to the remarkable downfield shift (-0.45 p.p.m.) of the signal peak of C_{18} -Me caused by C_{15} -BOH, the effect of C_{15} - α OH was as smaller as -0.10 p.p.m. (Table II). The configuration of the third hydroxyl group in V may thus be rather of alpha.

Hydroboration-oxidation 8 of methyl Δ^{14} -3 α ,12 α -dihydroxycholate, m.p. 89 $^\circ$ (free acid: m.p. 259 $^\circ$), derived from methyl ester of II by the method of K. Yamasaki et al. 9 , yielded the trihydroxy derivative which was identical with V giving the free acid III as a hydrolysate and the triketo ester VI on chromate oxidation.

Consequently, the chemical structure of the product III from I by the ferro-ascorbate system may reasonably be elucidated as $3\alpha,12\alpha,15\alpha$ -trihydroxycholanic acid.

Methyl ester	Site of hydroxyl group	С ₁₈ -Н (т)	С ₁₉ -Н (т)
Cholanate		9.39	9.08
Lithocholate	3α	9.38	9.09
Chenodeoxycholate	3α, 7α	9.32	9.04
Ursodeoxycholate	3α, 7β	9.34	9.07
Deoxycholate	3α , 12α	9.30	9.07
Cholate	3α, 7α, 12α	9.23	9.03
Compound V	3α , 12α , 15α	9.20	9.03

Table I. Chemical Shifts of Angular Methyl Protons

Spectra were taken with a Hitachi Model H-6013 (60 MC) spectrometer, in pyridine solution containing tetramethylsilane as an internal standard.

Table II.	Substituent Effect of Hydroxyl Group on the Chemical
	Shifts of Angular Methyl Protons

Site of hydroxyl group	Difference in p.p.m. from parent compound		
	с ₁₈ -н	С ₁₉ -Н	
3α	-0.01 (-0.01)	+0.01 (-0.01)	
7α	-0.06 (-0.06)	-0.05 (-0.09)	
7 ß	-0.04 (-0.02)	-0.02 (-0.01)	
12α	-0.08 (-0.12)	-0.02 (-0.03)	
15α	-0.10 (-0.07)	-0.04 (-0.03)	
15ß	- (-0.45)	- (-0.07)	

Numbers in the brackets are those reported.

REFERENCES

- 1. B. Matkovics, P. Pénzes and Gy. Göndös, Steroids, 5, 451 (1965).
- 2. G. A. Haslewood and V. M. Wooton, Biochem. J., 49, 67 (1951).
- 3. H. Budzikiewicz, C. Djerassi and D. H. Williams, "Structure Elucidation of Natural Products by Mass Spectrometry" Vol. 11., Holden-Dav, Inc., 1964, p 104.
- 4. a) H. Budzikiewicz and C. Djerassi, J. Am. Chem. Soc., 84, 1430 (1962).
 - b) P. Eneroth, B. Gordon and J. Sjövall, J. Lip. Res., 7, 524 (1966).
- 5, C. Djerassi, G. von Mutzenbecher, J. Fajkos, D. H. Williams and H. Budzikiewicz, J. Am. Chem. Soc., 87, 817 (1965).
- 6. C. Beard, J. M. Wilson, H. Budzikiewicz and C. Djerassi, J. Am. Chem. Soc., 86, 269 (1964).
- 7. a) K. Tori and K. Aono, Ann. Rept. Shionogi Res. Lab., 14, 136 (1964).
 - b) K. Tori, E. Kondo, Steroids, 4, 713 (1964).
- 8. a) H. C. Brown, "Hydroboration", W. A. Benjamin Inc. Publ., 1962.
 - b) G. Zweitov, N. R. Ayyanger and H. C. Brown, J. Am. Chem. Soc., 85, 2072 (1963).
- 9. K. Yamasaki, Z. Physiol. Chem., 220, 42 (1933); 233, 10 (1935).

Fig.1



